

The effects of fertilization regime on growth parameters and bioactive properties of pot grown *Cichorium spinosum* L. plants



Nikolaos Polyzos¹, Beatriz Paschoalinotto², Maria Compocholi¹, Maria Inês Dias², Lillian Barros², Spyridon A. Petropoulos^{1*}

¹University of Thessaly, Department of Agriculture, Crop Production and Rural Environment, Fytokou Street, 38446, Volos, Greece

²Centro de Investigação de Montanha (CIMO), Instituto Politécnico de Bragança, Campus de Santa Apolónia, 5300-253 Bragança, Portugal

*Corresponding author: spetropoulos@uth.gr



INTRODUCTION

- Cichorium spinosum* L. is a wild edible species, commonly known as stamnagathi found in many parts of Greece especially in Crete area and other Mediterranean countries.
- It is an integral part of the Mediterranean diet with people from rural communities usually hand picking the rosettes of the plant and use them in many traditional dishes.
- It is a plant that presents a wide adaptability that can be grown even coastal areas with low soil fertility, exhibits considerable tolerance to salt stress, while its cultivation demands are quite minimal regarding to its nutrition.
- Stamnagathi presents a high content in vitamins E (α - and γ -tocopherols) and K1, antioxidants, ω -3 fatty acids and several mineral elements; hence for that reason has been acknowledged as functional healthy food.
- The aim of the current study was to evaluate the effects of fertilization regime on growth parameters, chemical composition and bioactive properties of pot grown *C. spinosum* plants.

MATERIALS AND METHODS

- Young seedlings of *Cichorium spinosum* after emergence were transplanted in 2 L plastic pots containing peat and perlite (1:1, v/v).
- Seven treatments were used which varied in the amounts of N:P:K namely 100:100:100 (C111), 200:100:100 (C211), 200:200:200 (C222), 300:100:100 (C311), 300:200:200 (C322) and 300:300:300 (C333) ppm ratio of N:P:K and control (C0) where no fertilizers were added were applied via nutrient solution in the *C. spinosum* plants.
- Each treatment contained fifteen pots (n=15) and in total they were used 105 pots. All the treatments received the same amount of nutrient solution in which the plants were fertigated manually once a week comprising of 150 mL of N-P-K per plastic plot.
- Before harvest, it was recorded the chlorophyll content of leaves (SPAD index values) per treatment and after harvest the growth traits namely number of leaves/plant, weight of leaves/plant (g), dry matter of leaves (%), leaf area index (cm²) and specific leaf area index (m²/kg).
- Regarding the chemical composition and bioactive properties of the plants they were evaluated the identification and quantification of the phenolic compounds, antioxidant, anti-inflammatory, hepatotoxic and cytotoxic activities.



Image 1. The effects of fertilization regime on growth parameters and bioactive properties of pot grown *Cichorium spinosum* L. plants.

Table 1. The effects of fertilization regimes on number of leaves/plant, weight of leaves/plant (g), dry matter of leaves (%), chlorophyll content of leaves (SPAD index values), leaf area index (cm²) and specific leaf area index (m²/kg) of *C. spinosum* (Mean \pm SD).

Treatments	Traits					
	Number of leaves/plant	Weight of leaves/plant (g)	Dry matter (%)	SPAD index values	Leaf area index (cm ²)	Specific leaf area index (m ² /kg)
Co	29.54 \pm 1.35 (a)	11.49 \pm 0.97 (c)	8.27 \pm 2.16 (a)	94.81 \pm 12.21 (a)	297.12 \pm 8.50 (b)	27.09 \pm 1.69 (e)
C111	29.14 \pm 1.13 (a)	11.59 \pm 1.22 (c)	6.55 \pm 1.02 (e)	82.82 \pm 7.79 (bc)	282.82 \pm 7.86 (c)	31.17 \pm 1.71 (bc)
C211	24.21 \pm 1.38 (c)	12.90 \pm 1.35 (a)	6.69 \pm 0.09 (c)	74.19 \pm 6.61 (c)	324.75 \pm 8.57 (a)	28.20 \pm 1.73 (e)
C222	27.33 \pm 0.73 (b)	9.91 \pm 1.12 (e)	6.08 \pm 1.24 (e)	98.14 \pm 13.10 (a)	260.23 \pm 11.39 (d)	37.61 \pm 1.98 (a)
C311	27.29 \pm 1.27 (b)	12.02 \pm 1.69 (b)	5.56 \pm 2.55 (f)	62.10 \pm 7.00 (d)	250.42 \pm 6.76 (d)	39.20 \pm 1.45 (a)
C322	30.36 \pm 1.73 (a)	11.54 \pm 1.26 (c)	7.93 \pm 1.05 (b)	87.89 \pm 7.24 (ab)	278.37 \pm 8.28 (c)	30.82 \pm 1.64 (cd)
C333	29.73 \pm 1.30 (a)	10.85 \pm 0.77 (d)	6.92 \pm 2.57 (c)	76.08 \pm 7.79 (c)	255.81 \pm 7.99 (d)	29.78 \pm 1.46 (d)

*Mean values and standard deviations in the same column followed by different Latin letters are significantly different at $p < 0.05$ according to Student's t-test.

Table 2. Retention time (Rt), wavelength of the maximum absorption (λ max), deprotonated ion ([M-H]⁻), main mass fragments (MS²) and tentative identification of the phenolic compounds found in the hydroethanolic and aqueous extracts of *C. spinosum* samples.

Peak	Rt	λ max	[M-H] ⁻	MS ²	Tentative identification
1	8.51	292	337	191(100),173(12),163(71),155(3),119(34)	3-O-p-Coumaroylquinic acid
2	10.64	292	337	191(5),173(100),163(39),155(10),119(23)	4-O-p-Coumaroylquinic acid
3	17.24	352	477	301(100)	Quercetin-O-hexuronoside
4	17.82	345	461	285(100)	Luteolin-O- hexuronoside
5	18.97	342	505	463(24),301(100)	Quercetin-O-acetylhexoside
6	20.71	342	461	285(100)	Kaempferol-O- hexuronoside
7	22.07	340	491	315(100)	Isorhamnetin-O- hexuronoside
8	23.19	343	489	285(100)	Kaempferol-O-acetylhexoside
9	24.48	344	519	315(100)	Isorhamnetin-O-acetylhexoside

- The highest number of leaves was recorded by the C322 treatment, the highest fresh weight was noted for the C211 treatment and the highest SPAD index values was achieved by control treatment (C0).
- Related to leaf area index, the treatment C211 had the highest leaf area, while the C311 treatment recorded the highest specific leaf area index and the C0 treatment the lowest one respectively.
- In the case of hydroethanolic extracts, the highest antioxidant activity for the OxHLIA and TBARS assays was recorded by the C111 and C311 treatments, respectively, whereas the treatment C222 showed the best results for OxHLIA and TBARS assays in the aqueous extracts.
- Nine phenolic compounds were detected in both extracts, including two phenolic acids and seven flavonoids, with the major compounds being 4-O-p-coumaroylquinic acid and isorhamnetin-O-hexuronoside, regardless of the extraction method.
- The treatment of C333 contained the highest amounts of total phenolic acids, regardless of the extraction method. In the case of the hydroethanolic extracts, the highest content of total flavonoids was recorded by C311 treatment, total flavonoids, whereas regarding the aqueous extracts the highest content of total flavonoids was recorded by the C222 and C333 treatments
- The C311 treatment achieved the most content of total phenolic compounds in the case of the hydroethanolic extracts, whereas the C333 treatment recorded the highest content for the aqueous extracts
- Both extracts, did not show cytotoxic or anti-inflammatory activities.

RESULTS

Table 3. Antioxidant (IC₅₀ values μ g/mL), anti-inflammatory (IC₅₀ values μ g/mL), hepatotoxic (GI₅₀ values μ g/mL) and cytotoxic (GI₅₀ values μ g/mL) activities of the hydroethanolic extracts (HE) and aqueous extract (AE) of *C. spinosum* (Mean \pm SD).

Bioactive properties		Treatments						
		Co	C111	C211	C222	C311	C322	C333
Antioxidant	OxHLIA	322 \pm 20b	53 \pm 3g	339 \pm 18a	61 \pm 2f	65 \pm 2e	103 \pm 4d	123 \pm 7c
	Activity ^A - (HE/AE)	131 \pm 5c	97 \pm 5d	25 \pm 2e	20 \pm 1f	207 \pm 12b	278 \pm 9a	207 \pm 13b
Anti-inflammatory	RAW 264.7	>400	>400	>400	>400	>400	>400	>400
	Activity ^B - (HE/AE)	>400	>400	>400	>400	>400	>400	>400
Hepatotoxicity	PLP2	>400	>400	>400	>400	>400	>400	>400
	Activity ^C - (HE/AE)	>400	>400	>400	>400	>400	>400	>400
Cytotoxicity Activity ^D - (HE/AE)	AGS	>400	>400	>400	>400	>400	>400	>400
	CaCo2	>400	>400	>400	>400	>400	>400	>400
	VERO	>400	>400	>400	>400	>400	>400	>400
	MCF7	>400	>400	>400	>400	>400	>400	>400
		>400	>400	>400	>400	>400	>400	>400
		>400	>400	>400	>400	>400	>400	>400

^A Trolox IC₅₀ values: 5.8 \pm 0.6 μ g/mL (TBARS), 21.8 \pm 0.3 μ g/mL (OxHLIA 60 min); ^B Dexametaxone IC₅₀ value: 6.3 \pm 0.4 μ g/mL; ^C Ellipticine GI₅₀ values: 1.4 \pm 0.1 μ g/mL (PLP2), 1.23 \pm 0.03 μ g/mL (AGS), 1.21 \pm 0.02 μ g/mL (CaCo2), 1.41 \pm 0.06 μ g/mL (VERO) and 1.02 \pm 0.02 μ g/mL (MCF-7).

Table 4. Quantification (mg/g extract) of the phenolic compounds found in the hydroethanolic extracts (HE) and aqueous extracts (AE) of *C. spinosum* samples (Mean \pm SD).

Peak	Treatments						
	Co	C111	C211	C222	C311	C322	C333
1- (HE/EA)	0.54 \pm 0.003a	0.47 \pm 0.002c	0.45 \pm 0.003c	0.45 \pm 0.001c	0.51 \pm 0.003b	0.31 \pm 0.001d	0.46 \pm 0.002c
	0.46 \pm 0.000b	0.38 \pm 0.001c	0.37 \pm 0.004c	0.34 \pm 0.005d	0.44 \pm 0.003b	0.38 \pm 0.011c	0.52 \pm 0.01a
2- (HE/EA)	0.86 \pm 0.004cd	0.938 \pm 0.008b	0.81 \pm 0.001e	0.89 \pm 0.016c	0.84 \pm 0.004de	0.69 \pm 0.001f	1.01 \pm 0.005a
	0.57 \pm 0.006d	0.584 \pm 0.001c	0.59 \pm 0.001c	0.66 \pm 0.002b	0.65 \pm 0.014b	0.68 \pm 0.007b	0.82 \pm 0.001a
3- (HE/EA)	0.49 \pm 0.001b	0.46 \pm 0.000c	0.45 \pm 0.0001c	0.46 \pm 0.000c	0.53 \pm 0.000a	0.49 \pm 0.000b	0.48 \pm 0.002b
	0.53 \pm 0.002c	nd	0.47 \pm 0.001d	0.60 \pm 0.000a	0.53 \pm 0.001c	0.49 \pm 0.001d	0.56 \pm 0.001b
4- (HE/EA)	0.54 \pm 0.000c	0.53 \pm 0.000c	0.50 \pm 0.000d	0.49 \pm 0.000d	0.61 \pm 0.001v	0.53 \pm 0.001c	0.63 \pm 0.002a
	0.59 \pm 0.001c	0.52 \pm 0.000f	0.54 \pm 0.000e	0.63 \pm 0.001b	nd	0.56 \pm 0.001d	0.66 \pm 0.01a
TPA - (HE/EA)	1.40 \pm 0.002b	1.40 \pm 0.010b	1.27 \pm 0.002d	1.34 \pm 0.015c	1.36 \pm 0.006c	1.01 \pm 0.000e	1.48 \pm 0.002a
	1.03 \pm 0.006d	0.96 \pm 0.000f	0.96 \pm 0.003f	1.00 \pm 0.007e	1.10 \pm 0.018b	1.06 \pm 0.005c	1.34 \pm 0.009a
TF - (HE/EA)	3.67 \pm 0.002c	3.53 \pm 0.003d	3.41 \pm 0.001e	3.44 \pm 0.001e	3.97 \pm 0.000a	3.75 \pm 0.001b	3.75 \pm 0.001b
	3.03 \pm 0.008b	1.55 \pm 0.005e	2.81 \pm 0.003d	3.27 \pm 0.001a	3.04 \pm 0.010b	2.94 \pm 0.000c	3.27 \pm 0.008a
TPC - (HE/EA)	5.08 \pm 0.000c	4.94 \pm 0.013d	4.68 \pm 0.003f	4.79 \pm 0.015e	5.33 \pm 0.007a	4.76 \pm 0.001e	5.23 \pm 0.003b
	4.07 \pm 0.002d	2.52 \pm 0.005g	3.77 \pm 0.000f	4.27 \pm 0.008b	4.15 \pm 0.027c	4.01 \pm 0.005e	4.61 \pm 0.001a

nd - not detected. TPA - Total Phenolic Acids; TF - Total Flavonoids; TPC - Total Phenolic Compounds. Standard calibration curves used for quantification: p-coumaric acid ($y = 301.950x + 6966.7$, R² = 1, LOD = 0.68 μ g/mL and LOQ = 1.61 μ g/mL, peaks 1 and 2) and quercetin-3-O-glucoside ($y = 34.843x - 160.173$, R² = 0.9998, LOD = 0.21 μ g/mL; LOQ = 0.71 μ g/mL, peaks 3 to 9).

CONCLUSIONS AND RECOMMENDATIONS

- The application of fertilizers on *C. spinosum* plants had positive effects on plant growth, especially the 200:100:100 treatment where the highest fresh yield was recorded, while variable effects of fertilizer regimes on the chemical composition and bioactive properties were recorded.
- Discovering the optimal cultivation practices regarding the fertilizer management of spiny chicory could provide a promising outcome to the agronomic parameters, chemical composition and bioactive properties of the crop.
- Commercial cultivation of wild edible species is a promising cropping alternative in the climate change conditions as well in degraded soils where conventional crops cannot be cultivated or their yield is severely compromised.
- Further research is demanded in terms of evaluating the cultivation protocol in order to establish the commercial cultivation of the wild edible species.

REFERENCES

- Petropoulos, S. A., Fernandes, Á., Ntatsi, G., Levizou, E., Barros, L., & Ferreira, I. C. F. R. (2016). Nutritional profile and chemical composition of *Cichorium spinosum* ecotypes. *LWT - Food Science and Technology*, 73, 95–101. <https://doi.org/10.1016/j.lwt.2016.05.046>
- Petropoulos, S. A., Fernandes, Á., Tzortzakakis, N., Sokovic, M., Ciric, A., Barros, L., & Ferreira, I. C. F. R. (2019). Bioactive compounds content and antimicrobial activities of wild edible Asteraceae species of the Mediterranean flora under commercial cultivation conditions. *Food Research International*, 119(July 2018), 859–868. <https://doi.org/10.1016/j.foodres.2018.10.069>
- Petropoulos, S., Fernandes, Á., Karkanis, A., Antoniadis, V., Barros, L., & Ferreira, I. C. F. R. (2018). Nutrient solution composition and growing season affect yield and chemical composition of *Cichorium spinosum* plants. *Scientia Horticulturae*, 231(August 2017), 97–107. <https://doi.org/10.1016/j.scienta.2017.12.022>

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